Check for updates

The effect of physical exercise on anticancer immunity

Carmen Fiuza-Luces \mathbf{D}^{1} , Pedro L. Valenzuela^{1,2}, Beatriz G. Gálvez $\mathbf{D}^{1,3}$, Manuel Ramírez^{4,5,6}, Alejandro López-Soto^{7,8,9}, Richard J. Simpson^{10,11,12} & Alejandro Lucia $\mathbf{D}^{13,14}$

Abstract

Regular physical activity is associated with lower cancer incidence and mortality, as well as with a lower rate of tumour recurrence. The epidemiological evidence is supported by preclinical studies in animal models showing that regular exercise delays the progression of cancer, including highly aggressive malignancies. Although the mechanisms underlying the antitumorigenic effects of exercise remain to be defined, an improvement in cancer immunosurveillance is likely important, with different immune cell subtypes stimulated by exercise to infiltrate tumours. There is also evidence that immune cells from blood collected after an exercise bout could be used as adoptive cell therapy for cancer. In this Perspective, we address the importance of muscular activity for maintaining a healthy immune system and discuss the effects of a single bout of exercise (that is, 'acute' exercise) and those of 'regular' exercise (that is, repeated bouts) on anticancer immunity, including tumour infiltrates. We also address the postulated mechanisms and the clinical implications of this emerging area of research.

Physical Activity and Health Research Group (Pareig), Research institute of the Hospital 12 de Octubre
('imas12'), Madrid, Spain. ² Systems Biology Department, Universidad de Alcalá, Alcalá de Henares, Spain.
³ Department of Biochemistry and Molecular Biology, Faculty of Pharmacy, Universidad Complutense de Madrid,
Madrid, Spain. ⁴ Oncohematology Unit, Hospital Infantil Universitario Niño Jesús, Madrid, Spain. ⁵ Biomedical
Research Foundation, Hospital Infantil Universitario Niño Jesús, Madrid, Spain. ⁶ La Princesa Institute of Heah,
Madrid, Spain. ⁷ Department of Biochemistry and Molecular Biology, Faculty of Medicine, University of Oviedo,
Oviedo, Spain. ⁸ Instituto Universitario de Oncología del Principado de Asturias (IUOPA), Asturias, Spain. ⁹ Instituto
de Investigación Sanitaria del Principado de Asturias (ISPA), Asturias, Spain. ¹⁰ School of Nutritional Sciences
and Wellness, The University of Arizona, Tucson, AZ, USA. ¹¹ Department of Paediatrics, The University of Arizona,
Tucson, AZ, USA. ¹² Department of Immunobiology, The University of Arizona, Tucson, AZ, USA. ¹³ CIBER of Frailty
and Healthy Aging (CIBERFES), Madrid, Spain. ¹⁴ Faculty of Sport Sciences, Universidad Europea, Madrid, Spain.
🖂 e-mail: cfiuza.imas12@h12o.es; lopezsalejandro@uniovi.es; alejandro.lucia@universidadeuropea.es

and A stick and the shell Designed Owner ((Delland)). Designed to stick to shell a stick to shell a stick to she

▲ .*	
Sections	
0000113	

Introduction

Muscle tissue and immunity

Immune effects of acute and regular exercise

Effects on tumour immune cell infiltration

Conclusion

Introduction

The global burden of cancer – a leading cause of death worldwide – continues to grow, accounting for approximately 10 million deaths in 2020 (one in six of all deaths)¹. About half of all cancer deaths could be prevented by modifying lifestyle and environmental-related risk factors². Physical inactivity, which has reached pandemic proportions, is a modifiable risk factor that is gaining increasing attention². Indeed, nearly one in three adults worldwide³ fail to meet the minimum World Health Organization (WHO) recommendations for aerobic physical activity, which is defined as ≥150 min per week of moderate activities (such as walking) or \geq 75 min per week of vigorous activities (such as brisk or very brisk walking), or a combination thereof⁴. Physical inactivity starts early in life, with approximately 80% of children aged 11-17 years currently inactive⁵. In turn, regular physical activity is associated with lower cancer incidence⁶⁻⁸, lower cancer recurrence⁹ and reduced mortality⁸⁻¹¹. This association is independent of important confounders (such as body mass index or smoking status)⁶ and is potentially dose-response dependent, with a benefit threshold for mortality at approximately 3-5 times the minimum WHO dose and no excess risk at 10 times or more of the minimum dose¹¹.

The identification of the mechanisms underlying the potential antitumorigenic effects of exercise or physical activity, which remain largely unknown, will deepen our understanding of cancer biology. The stimulation of immune function is a strong candidate to at least partly explain these effects. The fact that the immune system is highly responsive to exercise-induced stimuli has been known for more than a century¹² (Box 1). The concept of 'exercise-induced leucocytosis', whereby a single bout of dynamic, moderate-vigorous (but not necessarily exhausting) exercise (typically running or bicycling for approximately 1 hour) elicits a profound mobilization of all major leucocyte subtypes into the peripheral circulation, was subsequently validated¹³⁻¹⁶. However, it was found that exercise-induced leucocytosis is not uniform and that immune cell subtypes with higher cytotoxicity, antigen experience and potential for migration into tissues (such as natural killer (NK) cells, v δ T cells and CD8⁺ T cells¹⁷, as well as CD14⁺CD16⁺ monocytes¹⁸ and CD16⁻ neutrophils¹⁹) are preferentially mobilized¹⁶. Moreover, there is evidence for a stimulating effect of exercise on immune effectors (particularly NK cells and CD8⁺ T cells) against tumours²⁰ and that optimal systemic immunity is critical for durable clinical responses to immunotherapy^{21,22}, as well as for prolonging the survival of patients with cancer in general²³.

Here we argue that the contracting muscle can be considered a modulator of immunity that can boost anticancer immune responses. We first discuss immune modulation by muscle tissue and the effects of acute and regular exercise (that is, a single exercise bout and repeated bouts, respectively) on the immune system, and then examine how the 'exercise milieu' can stimulate different immune cell subsets to infiltrate tumours and impair cancer growth. Throughout, we discuss the mechanisms - both at the systemic and tumour level - that may mediate these exercise effects and the potential clinical implications of the evidence available.

Muscle tissue and immunity

Skeletal muscle is an endocrine organ that releases a plethora of signalling molecules into the bloodstream. These include mainly proteins or small peptides (for example, cytokines, such as interleukin-6 (IL-6), IL-7 and IL-15), but also nucleic acids, lipids and metabolites (such as lactate). These can be freely circulating or packaged in exosomes and/or microvesicles and are collectively referred to as 'myokines'²⁴.

In addition to pleiotropic effects at the metabolic and multisystem level, myokines mediate immune-specific effects²⁵. For example, the contracting muscle releases IL-6, which rises exponentially with exercise intensity and duration, potentially reaching an approximately 100-fold increase over baseline circulating levels²⁶. Although IL-6 derived from other sources (such as immune cells) has a predominantly pro-inflammatory role, when released in the exercise milieu, it elicits an overall anti-inflammatory effect – notably, by inducing the release of other circulating cytokines that have anti-inflammatory properties, specifically IL-1 receptor antagonist (IL-1RA, a natural inhibitor of the potent pro-inflammatory cytokine IL-1\beta) and IL-10²⁷, while decreasing circulating levels of TNF²⁸. Additionally, beyond non-immune (mostly metabolic) functions²⁹, exercise-induced IL-6 can bind to NK cells and stimulate their acute mobilization to the bloodstream³⁰ and their homing into tumours³¹.

IL-7 and IL-15 are members of the common cytokine receptor y-chain family of cytokines and are also highly expressed or secreted by contracting muscles³²⁻³⁵. IL-7 helps to maintain thymic mass and promotes the output of naive T cells³⁶, and both cytokines have major roles in $\alpha\beta$ and $\gamma\delta$ T cell survival and in CD8⁺ T cell homoeostasis, to replenish (acutely or chronically) naive and memory subsets³⁶⁻⁴⁰. Notably, naive and memory CD8⁺T cells require IL-7Rα signalling and both IL-7Ra-mediated and IL-15-mediated signals, respectively, for proliferation in a lymphopenic host³⁷. Furthermore, in vitro treatment of naive T cells with low doses of IL-7 and IL-15, in combination with anti-CD3mediated and anti-CD28-mediated T cell receptor activation, results in the generation of cells resembling memory stem T cells, a subset of long-lived cells that can differentiate into virtually any type of T cell⁴¹. In turn, muscle-specific ablation of IL-15 in mice with a chronic viral infection lowers the levels of virus-specific CD8⁺ T cells and increases the levels of exhausted T cells that express inhibitory checkpoint receptors such as PD1, CD244, LAG3 and TIGIT³⁹. An increase in muscle mass (of note, muscle accretion is safely feasible with exercise in patients with cancer⁴²) has essentially the opposite effect to muscle-specific IL-15 ablation on CD8⁺ T cells³⁹. Stimulation of peripheral CD3⁺CD4⁻CD8⁻ T cells (a large proportion of which consists of γδ T cells) with IL-15 upregulates the expression of activating surface receptors such as NKG2D, NKp30 and NKp44, and increases the secretion of effectors such as IFN γ and soluble TRAIL, reflecting their enhanced tumouricidal activity⁴³. Importantly, IL-15-stimulated γδ T cells are being investigated as an alternative to NK cells and $\alpha\beta$ T cells in adoptive immunotherapy, owing to their ability to target tumour cells using both innate and T cell receptor-mediated mechanisms, their capacity to enhance antigen-specific T cell responses and their potential to be used in an autologous or allogeneic setting (for example, in acute myeloid leukaemia)⁴⁴. Preclinical models have shown that ex vivo expansion of CAR-19 T cells with IL-7 and IL-15 contributes to superior cell proliferation, effector functions, trafficking, survival and antitumour activity of these cells compared with CAR-19 T cells grown in IL-2⁴⁵. In patients with advanced-stage lymphoma who were treated with CAR-19 T cells, peak levels of IL-15 in the serum correlated with peak circulating levels of these cells and patients achieving remission showed higher levels of IL-15 than those who did not⁴⁶. Furthermore, the IL-15-IL-15Rα axis was shown to be involved in the infiltration of 'exercise-mobilized' CD8+ T cells in a mouse model of pancreatic cancer⁴⁷. This was reflected by a higher expression of genes downstream of IL-15 engagement in the CD8⁺ T cells of mice subjected to exercise training compared with their untrained controls, as well as by a significant increase in the fraction and number of IL-15R α -positive CD8 T⁺ cells in the tumours of

Box 1

Exercise, cancer and immunity: early observations

In 1921, Siverten and Dahlstrom postulated a prophylactic effect of 'muscular activity' against carcinoma¹⁴⁴. They based their theory on the observation that carcinoma incidence was higher in retired inactive farmers than in those remaining physically active until their seventies or eighties, as well as on the impression that carcinomas were rarely found in wildlife animals with high levels of spontaneous activity, such as rats. A proof-of-concept biological evidence for a protective association between physical exercise and cancer was reported in 1944 by Rush and Kline in albino mice that were inoculated with fibrosarcoma¹⁴⁵. Forced exercise – accomplished by rotating the cages continuously - starting 1 week before tumour inoculation and applied for 2 or 16 hours per day, delayed tumour growth rate by approximately 34% or 25%, respectively, compared with controls on an isocaloric diet¹⁴⁵. The potential for regular exercise to not only delay tumour growth (which was also shown in rat pups¹⁴⁶ or adult rats¹⁴⁷ implanted with a breast cancer cell line) but also reduce lung metastases in some mice injected

with tumorigenic cells (H-ras-transformed fibroblasts known as CIRAS-1 cells)¹⁴⁸ was essentially replicated over the following decades. However, whether exercise effects were linked to immune function was not contemplated despite the fact that both the exercise-induced leucocytosis phenomenon¹² and the phenomenon of immunosurveillance against tumours had already been documented. Virchow had identified in 1863 that neoplastic tissues were often infiltrated with leucocytes¹⁴⁹, and William Coley (often referred to as the 'the father of immunotherapy') had attempted to leverage the immune system to treat cancer in 1891^{150,151}. A study investigating whether the protective effect of exercise or physical activity against cancer was linked to an improved antitumorigenic function was published in 1993. Voluntary wheel running in mice for 9 weeks before the administration of CIRAS-1 cells increased splenic NK cell cytotoxic function for up to 3 weeks after the running intervention, although it did not impact tumour development¹⁵².

exercised mice, together with an upregulation of markers of proliferation and activation in tumour-infiltrating IL-15R α -positive CD8⁺T cells compared with their IL-15R α -negative counterparts⁴⁷.

The progressive decrease in myokine secretion owing to the ageing-related decline in skeletal muscle mass (sarcopenia) is a critical mechanism for the development of immunosenescence^{25,48}, a condition linked to cancer immune evasion⁴⁹. Here, tailored exercise – especially 'resistance training' – can attenuate sarcopenia even in patients of very advanced age (that is, over 85 years)⁵⁰.

The induction of muscle-derived cytokines through repeated bouts of exercise supports the maintenance of healthy immune effector cell populations²⁵ and promotes an overall anti-inflammatory milieu⁵¹. The latter is particularly important when considering that systemic, low-grade chronic inflammation - characterized by the activation of immune components that are often distinct from those engaged during acute immune response⁵² – is a hallmark of ageing (the so-called inflammageing phenomenon⁵³) and numerous chronic noncommunicable conditions, including cancer⁵². There is indeed meta-analytic evidence for an association between higher levels of chronic inflammation markers such as C-reactive protein and cancer risk⁵⁴. Indeed, pharmacological blockade of pro-inflammatory factors (such as IL-1ß) can reduce lung cancer incidence and mortality in individuals with a mean age above 60, with high levels of systemic inflammation at baseline55. At the mechanistic level, ageing-associated chronic inflammation favours oncogenesis not only by increasing cellular turnover53 but also, at least partly, by favouring the accumulation of immunosuppressive cell types in the tumour microenvironment, as documented in mouse models for myeloid-derived suppressor cells (MDSCs), type 2 macrophages (M2) and FOXP3⁺ regulatory T (T_{reg}) cells⁵⁶.

Advances in 'omics' technologies have led to the broad concept (beyond myokines) of 'exerkines' – which are signalling moieties that are released in response to exercise by several organs (not just muscle tissue) and exert their effects via endocrine, paracrine and/or autocrine pathways²⁴. Some exerkines influence immune function, particularly adrenaline, the 'fight or flight' hormone released during exertion⁵⁷.

Immune effects of acute and regular exercise Immune effects of acute exercise

In humans, acute dynamic exercise bouts lasting \geq 20–60 min induce a biphasic response in lymphocytes. The initial response is characterized by dramatic lymphocytosis that affects mainly NK cells, which increase several-fold above baseline levels in the blood^{58,59}. The most responsive NK cells are the mature KIR⁺ or NKG2A⁻ NK cells⁶⁰. CD8⁺ T cells and $v\delta$ T cells (which increase by approximately twofold⁶¹ and threefold¹⁷, respectively) are also mobilized in response to acute exertion. Acute exercise preferentially mobilizes subsets of CD8⁺ T cells⁶¹ and CD3⁺CD56⁺NK T-like cells⁶² that exhibit surface phenotypes associated with increased differentiation (for example, KLRG1⁺, CD57⁺ and CD28⁻) and gene expression programmes associated with antitumour activity, and also mobilizes CD14⁺CD16⁺ monocytes over classical CD14⁺CD16⁻ monocytes^{16,18,57,63}. Lymphocyte subtypes that are not typically involved in cytotoxicity (such as CD4⁺T cells and B cells) are recruited into the blood to a significantly lesser extent¹⁶. Within the T cell compartment, highly differentiated subsets of CD4⁺ and CD8⁺ T cells (such as effector memory (EM) and CD45RA⁺CCR7⁻ effector memory(EMRA)cells)arepreferentiallymobilized over their less differentiated counterparts (such as naive and central memory cells)¹⁶.

Lymphocyte mobilization during exertion is proportional to effort intensity and is driven by increased blood pressure and shear forces that cause demargination from the vascular and tissue reservoirs (the lung, liver and spleen), which boosts the number of leucocytes travelling in the main axial blood flow of the peripheral circulation^{16,64} (Fig. 1). Mobilization is also principally promoted by adrenaline stimulation of β_2 -adrenergic receptors on the surface of lymphocytes, leading to endothelial detachment and recirculation of lymphocytes into the bloodstream^{57,64-66}. The molecular mechanisms by which exercise and

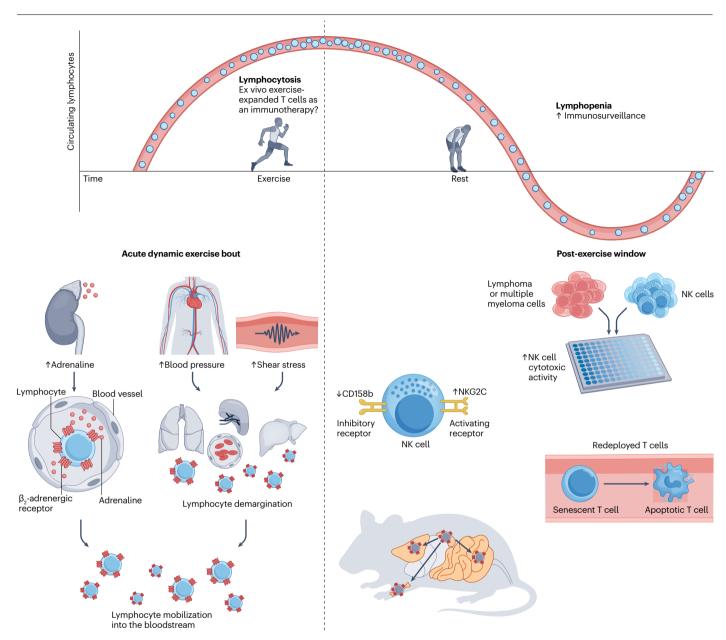


Fig. 1 | The biphasic immune cell response to dynamic (such as bicycling and running), acute exercise. Lymphocytosis – affecting mostly natural killer (NK) cells, CD8⁺ T cells and $\gamma\delta$ T cells – is driven by increased blood pressure and shear forces (causing demargination from the vascular or tissue reservoirs) and by adrenaline stimulation of β_2 -adrenergic receptors on the surface of lymphocytes. This is followed by transient (within 24 hours) lymphopenia where previously mobilized cells are redeployed to target tissues, with this post-exercise window

potentially improving immunosurveillance (higher NK cell cytotoxic capacity as assessed in vitro and decreased and increased proportion of NK cells expressing the inhibitory receptor CD158b and the activating receptor NKG2C, respectively). Because senescent T cells are mobilized into circulation to a greater extent than naive T cells, this could facilitate their apoptosis and create 'vacant space' for new naive T cells to take occupancy (see also Fig. 2).

catecholamine signalling affect immune cell mobilization, redistribution and function remain to be fully understood⁵⁷. However, shedding of the adhesion molecule ICAM-1 from the lymphocyte surface upon stimulation via adrenergic pathways might be one mechanism, with subsequent detachment of these cells from the vascular endothelium in primary (the bone marrow and thymus) and secondary (such as the spleen and lymph nodes) lymphocyte reservoirs⁶⁷, where norepinephrine and adrenaline are released from nerve terminals or diffuse from the bloodstream⁵⁷. In addition, there are indications that β -adrenergic stimulation can precipitate IL-6 release from skeletal muscle⁶⁸. The exertional lymphocyte mobilization response reflects, at least partly, the differential expression of β_2 -adrenergic receptors on lymphocyte subtypes – with the highest expression on NK cells, followed by $\gamma\delta$ T cells, CD8⁺ T cells, B cells and T_{reg} cells^{58,69} – and is

inhibited by non-selective β -antagonists that bind β_2 -receptors (nadolol and propranolol), but not by β_1 -only blockers (bisoprolol and metoprolol)^{18,65,70}.

Blood lymphocyte counts start to decrease during recovery after exercise, with a nadir at approximately 1-2 hours after exertion. Transient lymphopenia below pre-exercise levels is frequent, affecting mostly NK and CD8⁺T cells and gradually returning to baseline levels, usually within 24 hours⁵⁸. This acute, transient lymphopenia does not reflect immunosuppression and might occur in the context of an improved immunosurveillance⁵⁸. Indeed, in healthy individuals, NK cell cytotoxic capacity against lymphoma and multiple myeloma cell lines increases by 60% at 1 hour after exertion, which is accompanied by a decrease in the proportion of NK cells that express the inhibitory receptor CD158b and an increase in NK cells that express the activating receptor NKG2C⁶⁰. Fluorescent cell tracking studies in rodents revealed that T cells are largely redeployed from the spleen to target organs such as the lung, bone marrow and gut^{69,71}. Additionally, acute physical exercise preferentially mobilizes highly differentiated T cells into the circulation, many of which display phenotypes associated with exhaustion and terminal differentiation⁷². Some of these mobilized cells appear to be more susceptible to exercise-induced apoptosis, which may create 'vacant space' (especially if acute exertion bouts are repeated frequently) for new naive T cells to take occupancy⁷³. Through its ability to rapidly mobilize and increase circulating T cells, acute exercise has been postulated as a method to enrich T cells in the blood before leukapheresis, which will then be used for adoptive transfer immunotherapies that require ex vivo expansion (such as CAR T cells)²⁵ (Box 2).

Immune effects of regular exercise

The long-term beneficial effects of daily regular exercise might be due to the cumulative impact of 'repeated acute exercise bouts' and

the subsequent salutary effects during a few hours per day (Fig. 2). Since each bout of exercise induces myokine or exerkine secretion and induces the redeployment of massive numbers of lymphocytes, the effects of regular exercise on immune function in general and on antitumour immune function in particular might be linked, at least partly, to the progressive accumulation of frequent acute episodes of mobilization or redistribution of effector lymphocytes²⁵, even if this is not necessarily reflected by noticeable changes in the blood or in in vitro assessments performed under resting conditions in humans (that is, after a 'washout' period of \geq 24 hours from the last session). Immune adaptations to exercise training might be better observed using 'dynamic' endpoints such as the ability of NK cells or other immune cell populations to traffic to and infiltrate tumours²⁵.

Although a single exercise session mobilizes NK cells into the bloodstream, as corroborated for instance in patients with prostate cancer^{74,75}, this does not suffice to increase prostate NK cell infiltrates^{74,76}. Yet, a higher number of training sessions over an 8-week period correlated with greater prostate NK cell infiltrates in patients with this malignancy, and good adherence to the programme (up to 4 days a week) led to higher increases in tumour infiltrates at end intervention (mean change of +1.60 cells mm⁻²) compared with non-exercising controls (+0.44 cells mm⁻²)⁷⁷. This finding is relevant because prostate tumours hijack host macrophages to form a barrier around the tumour, preventing effector lymphocyte infiltration^{78,79}. Consequently, although a single exercise session does not change blood inflammatory markers in cancer survivors, there is a cumulative effect of subsequent sessions, with increases in IL-1RA⁸⁰.

In people with (or at risk of) cancer, exercise training interventions might not necessarily affect NK cell activity (as assessed in vitro with NK cells obtained from blood collected ≥ 1 day after the last exercise session)^{81,82}. There is, however, evidence for a significant enhancement in NK cell activity after an 8-week aerobic exercise intervention

Box 2

The use of exercise to facilitate the expansion of cells used for adoptive transfer

The use of 'exercise-expanded' T cells could overcome manufacturing issues such as the low cell numbers collected from lymphopenic patients, prolonged culturing to produce sufficient numbers of T cells or poor ex vivo cell expansion¹⁵³⁻¹⁵⁷. Adoptive transfer of ex vivo expanded tumour-associated antigen (TAA)specific cytotoxic T cells is a potential treatment strategy for several haematological and solid tumours and can reduce the risk of relapse following haematopoietic stem cell transplantation¹⁵⁷. Yet, this method can be hampered by difficulties in priming and expanding sufficient numbers of TAA-specific cytotoxic T cells²⁵. Here, the outcome of a study that collected blood lymphocytes from healthy adults at rest (baseline) and after a single bout of maximal-intensity exercise suggested that acute exercise might be used for immunotherapy with donor-derived T cells after allogeneic haematopoietic stem cell transplantation¹⁵⁸. TAA-specific cytotoxic T cells were expanded using autologous monocyte-derived dendritic cells pulsed with melanoma-associated antigen 4 (MAGE-A4), with preferentially expressed antigen in melanoma (PRAME) and with Wilms' tumour protein¹⁵⁸. Compared with baseline, post-exercise blood samples had a greater number of MAGE-A4-specific and PRAME-specific cells in 70% and 61% of participants (3.4-fold and 6.2-fold increase, respectively), and expanded TAA-specific cytotoxic T cells retained their antigen-specific cytotoxic activity¹⁵⁸. A similar approach was used to increase the expansion and antitumour activity of $\gamma\delta$ T cells, where cell products that were expanded from exercise-mobilized lymphocytes showed enhanced cytotoxic effects against a range of haematological tumours, which correlated with increases in the surface expression of activating receptors such as NKG2D⁷⁰. In summary, there is preliminary clinical evidence that immune cells from blood collected after an exercise bout could be used as adoptive T cell therapy for cancer.

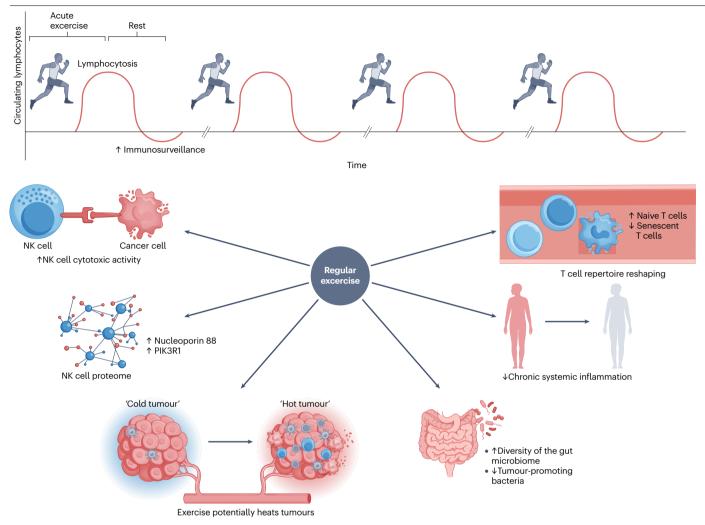


Fig. 2|**The immune cell response to regular exercise (that is, accumulation of repeated exercise bouts).** Regular exercise can increase natural killer (NK) cell cytotoxic activity (as assessed in vitro) against tumour cells, which can be accompanied by changes at the NK cell proteome level. Regular exercise might reshape the T cell repertoire by potentially increasing the proportion of naive

CD8⁺ T cells while also increasing immune cell infiltrates in tumours (see also Fig. 3). In addition, regular exercise attenuates chronic systemic inflammation – a condition associated with higher cancer risk – and increases gut microbiota diversity, while also potentially reducing the levels of tumour-promoting bacteria.

(including high-intensity interval training (HIIT)) in healthy adults⁸³, after a 12-week HIIT and resistance training intervention in patients with chronic lymphocytic leukaemia⁸⁴ and after an aerobic and resistance training programme performed during the first 9–12 weeks of (neo) adjuvant chemotherapy in women with resectable breast or colon cancer⁸⁵. Of note, training-induced increases in NK cell activity have been associated with changes at the proteome level, including the upregulation of PIK3R183, which is required for the maturation, homing, priming and function of NK cells⁸⁶, and nucleoporin 88 (ref. 83), which selectively mediates the nucleocytoplasmic transport of NF-KB, an ubiquitous transcription factor involved in immune responses⁸⁷. Regular exercise - or at least high aerobic fitness levels that usually result from exercise training – can also reshape the T cell repertoire by reducing the proportion of dysfunctional and senescent T cells that have an impaired ability to mediate antitumour responses²⁵, while increasing the proportion of naive CD8⁺ T cells (CD45RA⁺CD27⁺

 $CD62L^{+}CCR7^{+}$) that are capable of recognizing and responding to new antigens⁸⁸.

Beyond the direct effects of exercise on immune cells, a recent study that compared individuals with colorectal cancer who followed WHO guidelines of physical activity (179 participants, stages I–IV) with individuals who were inactive or obese has concluded that the beneficial effect of physical activity is associated with an enhancement of the alpha diversity (a measure of microbiome diversity) of the gut microbiota⁸⁹. Moreover, exercise also appeared to increase the abundance of bacteria that are associated with a retardation of tumour growth (such as *Faecalibacterium*) and reduced bacterial strains that have been associated with a promotion of tumour growth⁸⁹. The finding that higher microbial diversity is associated with regular physical activity or exercise is consistent with previous observations in athletes^{90,91} and is potentially important given that alpha diversity is a general indicator of good health⁹². Individuals with colorectal cancer have

previously been shown to have lower microbial diversity than healthy controls⁹³. In addition, there is evidence that the composition of the gut ecosystem markedly affects immune responses to cancer and the efficacy of antitumour immunotherapies^{94,95}. For example, a higher alpha diversity with a high abundance of *Faecalibacterium* has been associated with enhanced systemic and local antitumour immunity, mediated through increased antigen presentation and improved effector T cell function in the blood and the tumour microenvironment⁹⁶.

Effects on tumour immune cell infiltration

Several studies have specifically addressed the effects of exercise on the anticancer responses of different immune cell subsets. A large number of these studies have focused on NK cells, but others have also investigated exercise effects on myeloid cells, T cells and B cells.

NK cells

A seminal study led by Hojman showed that exposure of mice to regular voluntary wheel running (approximately 4–7 km per day from 4 weeks before to 2–3 weeks after tumour cell inoculation) reduced tumour incidence and growth by approximately 60% across five different tumour models (including B16-F10 melanoma)³¹. This was attributed, at least partly, to the combined effects of each of the repeated exercise bouts on catecholamine-induced mobilization of NK cells into the circulation and the subsequent binding of muscle-released IL-6 on NK cell surface receptors, ultimately driving NK cell homing to tumours³¹. Here, the amount of infiltrates was inversely linked to tumour burden, and NK cell depletion with antibodies abolished the suppressive effect of exercise on tumour growth³¹ (Fig. 3).

Another study confirmed higher tumour NK cell infiltrates in mice that performed forced treadmill running for 20 days before being orthotopically transplanted with breast cancer cells compared with control mice⁹⁷. Although running might not necessarily enhance NK cell cytotoxicity in mice per se, it might enhance NK cell infiltration into the tumour environment by upregulating the expression of genes encoding chemokines (such as Ccl3, Cxcl10 and Cx3cl1) and increase tumour immunogenicity via the upregulation of ligands for the NK cell-activating receptor NKG2D (such as *Mult1* and *H60a*)³¹. In a study where athymic mice (which lack functional T cells but retain NK cells) were challenged with prostate cancer cells and then treated with radiotherapy, a 2-week forced treadmill running intervention stimulated antitumour immune responses by enhancing NK cell tumour infiltration and upregulated gene expression of receptors required for NK cell cytotoxic activity (Klrk1, which encodes NKG2D, and Il2rb)98. In a recent proof-of-concept randomized controlled trial in the setting of preoperative prostatectomy, where patients were divided into a control (standard of care) or 8-week HIIT intervention group, NK cell infiltrates in prostatic tissue were analysed in diagnostic core needle biopsies before the intervention and upon prostatectomy (post-intervention)⁷⁷. This revealed increased NK cell infiltration in the exercise group compared to the control group⁷⁷. These findings are likely relevant when considering the important role that NK cells have in antitumour immunity⁹⁹⁻¹⁰², and that increased NK cell infiltrates inside tumours are associated with higher response to chemotherapy¹⁰³ and higher survival¹⁰³ in gastric cancer, and also with improved response to anti-PD1 immunotherapy treatment¹⁰⁴ and survival in metastatic melanoma105.

NK cells have been traditionally viewed as the main effectors of the exercise-related immunosurveillance against tumour development,

partly because they are the immune cell subset that is the most responsive to the acute mobilizing effect of a single exercise bout and thus have been the most studied. However, this may not always be the case. For example, a study of mice with B16-F10 melanoma has found that when the exercise intervention started after the implantation of the tumour. no changes in intratumoural NK cell infiltrates were observed¹⁰⁶. This stands in contrast to the findings by Hojman's group as discussed above and might suggest that a 'pre-conditioning' exercise effect before tumour onset may be needed to enhance NK cell infiltration. On the other hand, the study by Hojman's group reported larger tumours in athymic mice than in wild-type mice³¹, indicating that T cells had a major role (potentially beyond NK cells) in controlling tumour growth independently of exercise state and that all immune cells must work together to orchestrate an efficient antitumour response. Although there is overall preclinical and clinical evidence that regular exercise can increase tumour NK cell infiltration, whether this translates into actual clinical benefits remains to be determined.

Myeloid cells

Studies of the effects of exercise on myeloid anticancer responses have mainly focused on MDSCs, the polarization of tumour-associated macrophages (TAMs) and tumour infiltrates of dendritic cells. For example, two studies have reported that exercise training in a mouse model of breast cancer reduced the tumour-induced accumulation of MDSCs after 6 weeks of voluntary wheel running or approximately 3 weeks of treadmill running starting at different timepoints after the inoculation with cancer cells^{107,108}. This is potentially important because MDSCs (especially CD14⁺HLA-DR^{lo/neg} monocytes) can have a role in cancer-induced immunosuppression, notably by inhibiting the antitumour activities of T cells and NK cells at the same time as stimulating T_{reg} cells¹⁰⁹⁻¹¹¹. The reduction of intratumoural MDSC infiltrates after exercise training in mice was accompanied by a relative increase in NK and CD8⁺T cell activation and improved responses to combination therapy with PD1 blockade and radiotherapy¹⁰⁸. There is also evidence for an improved response to PD1 blockade when treadmill running was combined with resistance training in a patient-derived xenograft mouse model of lung cancer, albeit with no significant differences in total tumour MDSC infiltrates¹¹². Another study in a mouse model of melanoma (B16), however, has found no additive effect of exercise (wheel running) on the efficacy of immunotherapy (PD-1 or PD-L1 blockade)¹¹³.

Macrophages can be polarized towards a pro-inflammatory (M1) phenotype or an immunosuppressive (M2) phenotype. M1-polarized TAMs induced by IFNy and lipopolysaccharide are considered antitumorigenic owing to the expression of inducible nitric oxide synthase (iNOS) and the secretion of cytotoxic reactive oxygen species and pro-inflammatory cytokines¹¹⁴. By contrast, TAMs activated by IL-4 (M2-TAMs) can be considered pro-tumorigenic owing to their expression of growth-promoting, proangiogenic and extracellular matrix remodelling signals via vascular endothelial factor, among other factors¹¹⁴. M2-TAMs also show elevated expression of CCL22 (also known as monocyte chemoattractant protein-1)¹¹⁵ which facilitates the recruitment of immunosuppressive T_{reg} cells¹¹⁶. In a mouse model of invasive breast cancer, wheel-running training (especially above 150 km per week) had a dose-response effect on tumour growth, together with a significant reduction in Ccl22 transcription¹¹⁷. A study in a mouse model of intestinal tumorigenesis has found that, compared with sedentary controls, mucosal scrapings of treadmill-exercised mice (1 hour per day, 6 days per week for 12 weeks) had lower levels of transcripts of Ccl22, as well as of Cd206 and Arg1 - two other genes

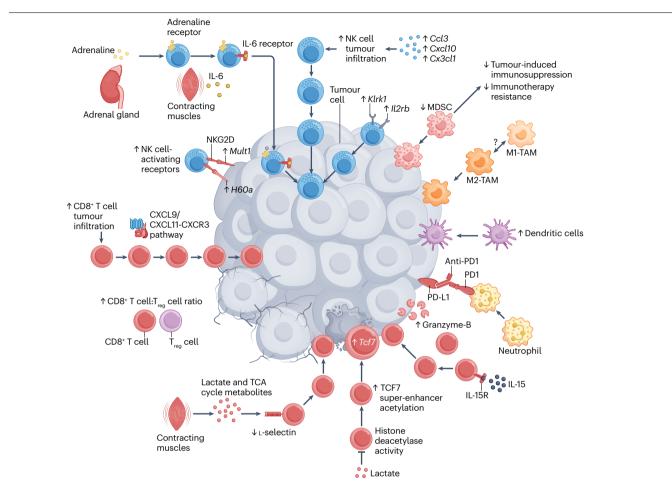


Fig. 3 | Regular exercise has the potential to 'heat' tumours. Exercise-primed immune cell infiltrates in tumours have mainly been studied with regard to natural killer (NK) cells, with the evidence indicating enhanced gene expression of chemokines that stimulate NK cell infiltration into the tumour bed (*Ccl3, Cxcl10* and *Cx3cl1*), increased tumour expression of ligands (*H60a, Mult1*) for the NK cell-activating receptor NKG2D, and gene upregulation of receptors required for NK cell cytotoxic activity (*Klrk1*, encoding NKG2D, and *Il2rb*). In addition, during each exercise bout, β_2 -adrenergic-driven NK cell mobilization into the blood is followed by myokine (IL-6)-mediated infiltration into tumours. Exercise training can reduce tumour infiltrates of an immune subset that has been linked to immunotherapy resistance and tumour progression: the myeloid-derived suppressor cells (MDSCs). There is more controversy regarding the potential infiltration by tumour-associated macrophages (TAMs). Regular exercise can also

increase tumour infiltrates of dendritic cells (this was shown in a mouse model of high-risk neuroblastoma, one of the most aggressive paediatric tumours¹²¹). There is evidence for a CXCL9–CXCR3 and CXCL11–CXCR3 pathway-mediated increase in tumour infiltrates – and effector functions – of CD8⁺T cells with exercise training, together with an inhibition of FOXP3⁺ regulatory T (T_{reg}) cells. Some myokines might prime CD8⁺T cells against tumours by decreasing the expression of the cell adhesion molecule L-selectin (lactate and tricarboxylic acid (TCA) metabolites), by epigenetic mechanisms (lactate released from muscles can inhibit histone deacetylase in T cells, which, in turn, leads to higher acetylation of H3K27 of the *Tcf7* super enhancer locus, leading to increased *Tcf7* gene expression) or by stimulating the release of granzyme-B (IL-15). Finally, neutrophils can infiltrate lung tumours in response to exercise training, with these cells acting as potential effector cells of PD1 inhibition.

that are upregulated in M2-TAMs – together with lower expression of FOXP3, a marker for T_{reg} cells¹¹⁸. By contrast, transcriptome analysis in a mouse model of lung cancer showed that a 12-week endurance (treamill exercise) regimen decreased the proportion of M1-TAMs (that is, lower mRNA levels of the M1 markers *Cd86*, *Tnf* and *Nos2* compared with controls)¹¹⁹. In a similar vein, other studies of breast tumour transcriptome analyses¹²⁰ or flow cytometry of high-risk neuroblastoma tumours¹²¹ indicated that wheel running in the 2 weeks before tumour engraftment¹²⁰ or treadmill running combined with resistance exercise for 5 weeks after tumour implantation¹²¹ led to higher M2-like TAM infiltrates in the tumours as compared with control mice.

However, the inhibitory effect of exercise on tumour growth might be independent of its effect on total TAM infiltrates, as antitumorigenic immune activity can occur on a background of decreasing levels of TAM infiltration¹¹⁹. Consequently, TAMs extracted from exercised lymphoma-bearing mice had a higher antitumour activity in vitro and production of tumouricidal molecules (such as TNF and nitric oxide) than those from non-exercised controls¹²².

Dendritic cells stimulate and expand tumour-specific effector T cells through IL-12 signalling, and a high number of dendritic cells are detected in spontaneous regressing tumour models, suggesting a critical role in tumour control^{123,124}. A 5-week treadmill running

and resistance intervention in a mouse model of high-risk neuroblastoma increased the presence of myeloid cells, and in particular dendritic cells, in tumours¹²¹. This finding is potentially relevant since the high-risk neuroblastoma tumour environment is often referred to as 'cold' or 'immune-deserted'¹²⁵ and strategies to increase dendritic cells represent a potential immunotherapeutic approach to promote tumour regression¹²⁶. Moreover, an 8-week moderate-intensity training regimen (treadmill aerobic exercise and resistance training) in a patient-derived xenograft mouse model of early-stage non-smallcell lung carcinoma decreased the tumour growth rate compared to controls, and also tended to increase intratumoural neutrophil infiltrates (that is, quasi-significant P value of 0.060) when combined with immunotherapy (PD1 blockade)¹¹². This finding is also of potential interest given that, although myeloid subsets have been less studied than lymphocytes in the context of immunotherapy, neutrophil infiltrates might exert a cytotoxic effect and act as effector cells (involved in necrotic tumor regression) in response to PD1 inhibition¹²⁷.

T cells

An 8-week forced running intervention in a mouse model of breast cancer slowed tumour growth and increased the ratio of intratumoural CD8⁺ T cells to T_{reg} cells¹²⁸. Similarly, exercise training for 1–2 weeks delayed the growth of breast cancer in an orthotopic rodent model by increasing tumour infiltration and effector functions of CD8+ T cells via CXCL9-CXCR3-mediated and CXCL11-CXCR3-mediated pathways¹²⁹. Beyond the finding that a delay in the growth of breast cancer tumours was observed in exercised wild-type mice but not in equivalent athymic mice¹²⁸, there is additional mechanistic evidence supporting a CD8⁺ T cell-dependent reduction in tumour growth in response to exercise: skeletal muscle-derived lactate and tricarboxylic acid cycle metabolites, which are released into circulation during exertion (and reach virtually all tissues including lymphoid organs), appear to enhance the antitumour effector profile of CD8⁺ T cells, notably by driving the loss of the cell adhesion molecule CD62L (also known as L-selectin)¹³⁰. Similarly, the administration of sodium lactate for 2-3 weeks in mice inoculated with MC-38 colon adenocarcinoma cells resulted in an immune-dependent decline in tumour expansion by enhancing the stemness of CD8⁺ T cells through epigenetic regulation; specifically, lactate inhibited histone deacetylase activity, resulting in increased acetylation (at H3K27) of a T cell-specific transcription factor (Tcf7) super enhancer locus, leading to increased Tcf7 gene expression¹³¹. Furthermore, there is recent evidence that lactate represents a bioenergetic and biosynthetic fuel for CD8⁺ effector T cells and also has an effect on their transcriptome, including the expression of key effector differentiation markers such as granzyme B (a serine protease secreted by these cells to mediate apoptosis in target cells) and IFNy^{132,133}.

Pancreatic ductal adenocarcinoma (PDA) is the third leading cause of cancer-related death in the USA with 5-year survival of <10% and no effective treatment for advanced disease^{134,135}. Furthermore, PDA remains resistant to immunotherapies, probably because of the modest T cell infiltrate and highly immunosuppressive tumour microenvironment^{134,135}. Some studies have found no changes with exercise in the tumour infiltrates of mice that were subcutaneously inoculated with PDA cells¹³⁶. Yet, a notable recent study has shown that 6-week treadmill running restricted tumour growth in several different mouse models of PDA, which was accompanied at the intratumoural level by a significant expansion of lymphocyte clusters, particularly CD8⁺T cells, and a contraction of MDSCs⁴⁷. The authors have noted that

only CD8⁺ T cells induced by exercise, particularly those responsive to IL-15 signalling, were responsible for the observed reductions in tumour growth⁴⁷. In addition, individuals with PDA who participated in a preoperative exercise training programme showed significantly higher tumour CD8⁺ T cell infiltrates and a trend towards higher expression of granzyme-B when compared with matched historical controls, in which higher levels of these infiltrates or granzyme-B were positively associated with survival in the exercise cohort⁴⁷. This study has provided mechanistic evidence for an exercise-primed immune effect and biological support for novel therapeutic targets, such as IL-15 signalling. Indeed, treatment with an IL-15 superagonist enhanced the efficacy of immunotherapy (PD1 blockade) and promoted durable responses in mice⁴⁷.

B cells

The concept of tumour-infiltrating lymphocytes (TILs) has evolved to include not only T or NK cells but also B cells and plasma cells, collectively referred to as TIL-Bs¹³⁷. TIL-Bs can perform important functions in the tumour microenvironment, including antigen presentation and antibody production, to support both T cell responses and innate immune mechanisms^{137,138}. The presence of high B cell infiltration inside tumours has been associated with improved response to immunotherapy and longer survival in patients with melanoma and sarcoma^{139,140}. However, TIL-Bs have rarely been studied in the context of exercise, with no significant differences reported in B cell infiltrates between exercised and control mice in models of high-risk neuroblastoma, PDA or several other tumour types^{31,47,121}. More research is, therefore, needed on this topic.

Cancer progression despite immune cell infiltration

The findings of exercise-induced tumour infiltrates are promising. Nevertheless, it is important to note that, although tumours can be recognized by the immune system (which can delay their growth before they become clinically apparent), cancer can still develop¹⁴¹. Indeed, tumour cells can progressively change their phenotype through genetic and epigenetic adaptations to escape from immunosurveillance and advance to a clinically detectable state either by 'hiding' from immune recognition ('immunoselection', also known as 'immunoediting', a process where non-immunogenic tumour cell variants have a selective advantage) or actively suppressing immune effectors ('immunosubversion')⁴⁹. In this context, although tumours in humans are often infiltrated by TILs, they may not exert significant antitumour activity¹⁴¹. Thus, it is conceivable that increasing the numbers of TILs alone does not necessarily allow for meaningful immune responses.

Glossary

Exercise (or 'exercise training')

A form of structured leisure-time physical activity with the purpose of improving or maintaining health training for a 10-km running race, or resistance training (for example, weight lifting) to increase muscle mass. Although physical activity and exercise are often used interchangeably, the bulk of observational epidemiological evidence is based on physical activity data, whereas exercise is frequently used in intervention trials and preclinical studies.

Physical activity

Any bodily movement produced by skeletal muscles that requires energy expenditure and includes the domains of occupational, domestic, transportation and leisure time (such as walking to work or walking the dog).

An increase in TILs mediated by exercise in combination with immune checkpoint inhibitor therapy might, however, help to attenuate tumour growth – at least theoretically.

Conclusion

There is biological evidence for an immune-stimulating effect of regular physical activity or exercise, notably, by stimulating immune cell mobilization (and, at least potentially, homing into tumours) in the few hours after each acute bout of exercise. As opposed to immunotherapeutic approaches, the beneficial immune effects of exercise are not accompanied by detrimental side effects, and carefully adapting exercise programmes to the individual characteristics of each patient can have a positive impact on health status, even in those with advanced-stage cancer¹⁴². These observations support the recommendation of experts that 'all people living with and beyond cancer should be as active as is possible for them¹⁴³. More research is, however, needed for testing the 'bench-to-bed' translation of recent preclinical findings, as well as new designs to unveil the potential mechanisms behind the preventive benefits of exercise against tumour establishment (that is before a tumour is already visible and assessable). In addition to expanding preclinical research, it would seem necessary to assess potential (co)adjuvant exercise effects in the context of immunotherapy trials and in the prehabilitation setting (that is, preoperative interventions, such as exercise training, aimed at increasing the physiological reserve of patients with cancer so that they can better withstand the stress of surgery on body systems). Some other questions regarding specific exercise modalities remain open, such as the potential effects of resistance exercise (which has been scarcely studied compared with endurance exercise) on anticancer immune function.

Published online: 04 October 2023

References

- Sung, H. et al. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 71, 209–249 (2021).
- GBD 2019 Cancer Risk Factors Collaborators. The global burden of cancer attributable to risk factors, 2010–19: a systematic analysis for the Global Burden of Disease Study 2019. Lancet 400, 563–591 (2022).
- Guthold, R., Stevens, G. A., Riley, L. M. & Bull, F. C. Worldwide trends in insufficient physical activity from 2001 to 2016: a pooled analysis of 358 population-based surveys with 1.9 million participants. *Lancet Glob. Health* 6, e1077–e1086 (2018).
- Bull, F. C. et al. World Health Organization 2020 guidelines on physical activity and sedentary behaviour. Br. J. Sports Med. 54, 1451–1462 (2020).
- Garcia-Hermoso, A. et al. Adherence to aerobic and muscle-strengthening activities guidelines: a systematic review and meta-analysis of 3.3 million participants across 32 countries. Br. J. Sports Med. 57, 225–229 (2023).
- Moore, S. C. et al. Association of leisure-time physical activity with risk of 26 types of cancer in 1.44 million adults. JAMA Intern. Med. 176, 816–825 (2016).
- Matthews, C. E. et al. Amount and intensity of leisure-time physical activity and lower cancer risk. J. Clin. Oncol. 38, 686–697 (2020).
- Ahmadi, M. N. et al. Vigorous physical activity, incident heart disease, and cancer: how little is enough? *Eur. Heart J.* 43, 4801–4814 (2022).
- Morishita, S. et al. Effect of exercise on mortality and recurrence in patients with cancer: a systematic review and meta-analysis. *Integr. Cancer Ther.* 19, 1534735420917462 (2020).
- Friedenreich, C. M., Stone, C. R., Cheung, W. Y. & Hayes, S. C. Physical activity and mortality in cancer survivors: a systematic review and meta-analysis. *JNCI Cancer Spectr.* 4, pkz080 (2019).
- Arem, H. et al. Leisure time physical activity and mortality: a detailed pooled analysis of the dose-response relationship. JAMA Intern. Med. 175, 959–967 (2015).
- 12. Larrabee, R. C. Leucocytosis after violent exercise. J. Med. Res. 7, 76–82 (1902).
- Nieman, D. C. & Wentz, L. M. The compelling link between physical activity and the body's defense system. J. Sport Health Sci. 8, 201–217 (2019).
- Simpson, R. J., Bigley, A. B., Agha, N., Hanley, P. J. & Bollard, C. M. Mobilizing immune cells with exercise for cancer immunotherapy. *Exerc. Sport Sci. Rev.* 45, 163–172 (2017).
- Simpson, R. J. et al. Human cytomegalovirus infection and the immune response to exercise. Exerc. Immunol. Rev. 22, 8–27 (2016).

- Simpson, R. J., Kunz, H., Agha, N. & Graff, R. Exercise and the regulation of immune functions. Prog. Mol. Biol. Transl. Sci. 135, 355–380 (2015).
- Anane, L. H. et al. Mobilization of gammadelta T lymphocytes in response to psychological stress, exercise, and beta-agonist infusion. *Brain Behav. Immun.* 23, 823–829 (2009).
- Steppich, B. et al. Selective mobilization of CD14(+)CD16(+) monocytes by exercise. Am. J. Physiol. Cell Physiol. 279, C578–C586 (2000).
- Peake, J. et al. Changes in neutrophil surface receptor expression, degranulation, and respiratory burst activity after moderate- and high-intensity exercise. J. Appl. Physiol. 97, 612–618 (2004).
- Fiuza-Luces, C., Valenzuela, P. L., Castillo-García, A. & Lucia, A. Exercise benefits meet cancer immunosurveillance: implications for immunotherapy. *Trends Cancer* 7, 91–93 (2021).
- Zuazo, M. et al. Functional systemic CD4 immunity is required for clinical responses to PD-L1/PD-1 blockade therapy. EMBO Mol. Med. 11, e10293 (2019).
- Spitzer, M. H. et al. Systemic immunity is required for effective cancer immunotherapy. Cell 168, 487–502 (2017).
- Gustafson, M. P. et al. Immune monitoring using the predictive power of immune profiles. J. Immunother. Cancer 1, 7 (2013).
- Chow, L. S. et al. Exerkines in health, resilience and disease. Nat. Rev. Endocrinol. 18, 273–289 (2022).
- Gustafson, M. P. et al. Exercise and the immune system: taking steps to improve responses to cancer immunotherapy. J. Immunother. Cancer 9, e001872 (2021).
- Fischer, C. P. Interleukin-6 in acute exercise and training: what is the biological relevance? Exerc. Immunol. Rev. 12, 6–33 (2006).
- Steensberg, A., Fischer, C. P., Keller, C., Moller, K. & Pedersen, B. K. IL-6 enhances plasma IL-1RA, IL-10, and cortisol in humans. *Am. J. Physiol. Endocrinol. Metab.* 285, E433–E437 (2003).
- Starkie, R., Ostrowski, S. R., Jauffred, S., Febbraio, M. & Pedersen, B. K. Exercise and IL-6 infusion inhibit endotoxin-induced TNF-α production in humans. *FASEB J.* 17, 884–886 (2003).
- Pedersen, B. K. & Febbraio, M. A. Muscle as an endocrine organ: focus on muscle-derived interleukin-6. *Physiol. Rev.* 88, 1379–1406 (2008).
- Bay, M. L. et al. Human immune cell mobilization during exercise: effect of IL-6 receptor blockade. Exp. Physiol. 105, 2086–2098 (2020).
- Pedersen, L. et al. Voluntary running suppresses tumor growth through epinephrine- and IL-6-dependent NK cell mobilization and redistribution. *Cell Metab.* 23, 554–562 (2016).

This elegant, mechanistic study shows the involvement of two exerkines, epinephrine and IL-6, on NK-cell mobilization into several tumour types.

- Quinn, L. S., Haugk, K. L. & Grabstein, K. H. Interleukin-15: a novel anabolic cytokine for skeletal muscle. *Endocrinology* 136, 3669–3672 (1995).
- Haugen, F. et al. IL-7 is expressed and secreted by human skeletal muscle cells. Am. J. Physiol. Cell Physiol. 298, C807–C816 (2010).
- Nielsen, A. R. et al. Expression of interleukin-15 in human skeletal muscle effect of exercise and muscle fibre type composition. J. Physiol. 584, 305–312 (2007).
- Tamura, Y. et al. Upregulation of circulating IL-15 by treadmill running in healthy individuals: is IL-15 an endocrine mediator of the beneficial effects of endurance exercise? Endocr. J. 58, 211–225 (2011).
- Capitini, C. M., Chisti, A. A. & Mackall, C. L. Modulating T-cell homeostasis with IL-7: preclinical and clinical studies. J. Intern. Med. 266, 141–153 (2009).
- Fry, T. J. & Mackall, C. L. The many faces of IL-7: from lymphopoiesis to peripheral T cell maintenance. J. Immunol. 174, 6571–6576 (2005).
- Goldrath, A. W. et al. Cytokine requirements for acute and basal homeostatic proliferation of naïve and memory CD8+ T cells. J. Exp. Med. 195, 1515–1522 (2002).
- Wu, J. et al. Skeletal muscle antagonizes antiviral CD8+T cell exhaustion. Sci. Adv. 6, eaba3458 (2020).

This study provides a mechanistic link between two seemingly isolated events that are prevalent in the context of cancer — loss of muscle mass and T cell exhaustion — and shows the importance of skeletal muscle preservation to protect the proliferative potential of these cells.

- Wallace, D. L. et al. Prolonged exposure of naïve CD8+ T cells to interleukin-7 or interleukin-15 stimulates proliferation without differentiation or loss of telomere length. *Immunology* 119, 243–253 (2006).
- Cieri, N. et al. Il-7 and IL-15 instruct the generation of human memory stem T cells from naive precursors. *Blood* 121, 573–584 (2013).
- Campbell, K. L. et al. Exercise guidelines for cancer survivors: consensus statement from international multidisciplinary roundtable. *Med. Sci. Sports Exerc.* 51, 2375–2390 (2019).
- 43. Yao, J. et al. Human double negative T cells target lung cancer via ligand-dependent mechanisms that can be enhanced by IL-15. *J. Immunother. Cancer* **7**, 17 (2019).
- Van Acker, H. H. et al. Interleukin-15 enhances the proliferation, stimulatory phenotype, and antitumor effector functions of human gamma delta T cells. J. Hematol. Oncol. 9, 101 (2016).
- Xu, Y. et al. Closely related T-memory stem cells correlate with in vivo expansion of CAR. CD19-T cells and are preserved by IL-7 and IL-15. Blood **123**, 3750–3759 (2014).
- Kochenderfer, J. N. et al. Lymphoma remissions caused by anti-CD19 chimeric antigen receptor T cells are associated with high serum interleukin-15 levels. J. Clin. Oncol. 35, 1803–1813 (2017).

- Kurz, E. et al. Exercise-induced engagement of the IL-15/IL-15Rα axis promotes anti-tumor immunity in pancreatic cancer. Cancer Cell 40, 720–737.e5 (2022).
 This mechanistic preclinical and clinical study highlights the therapeutic potential
- of regular exercise against one of the deadliest malignancies, pancreatic ductal adenocarcinoma, through IL-15-mediated activation of CD8' T cells.
 Nelke, C., Dziewas, R., Minnerup, J., Meuth, S. G. & Ruck, T. Skeletal muscle as potential
- central link between sarcopenia and immune senescence. *eBioMedicine* **49**, 381–388 (2019).
- López-Otín, C., Pietrocola, F., Roiz-Valle, D., Galluzzi, L. & Kroemer, G. Meta-hallmarks of aging and cancer. Cell Metab. 35, 12–35 (2023).
- Izquierdo, M., Morley, J. E. & Lucia, A. Exercise in people over 85. *BMJ* 368, m402 (2020).
 Fiuza-Luces, C. et al. Exercise benefits in cardiovascular disease: beyond attenuation of traditional risk factors. *Nat. Rev. Cardiol.* 15, 731–743 (2018).
- Furman, D. et al. Chronic inflammation in the etiology of disease across the life span. Nat. Med. 25, 1822–1832 (2019).
- López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M. & Kroemer, G. Hallmarks of aging: an expanding universe. *Cell* 186, 243–278 (2023).
- Michels, N., van Aart, C., Morisse, J., Mullee, A. & Huybrechts, I. Chronic inflammation towards cancer incidence: a systematic review and meta-analysis of epidemiological studies. *Crit. Rev. Oncol. Hematol.* **157**, 103177 (2021).
- Ridker, P. M. et al. Effect of interleukin-1β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet* **390**, 1833–1842 (2017).
- Fane, M. & Weeraratna, A. T. How the ageing microenvironment influences tumour progression. Nat. Rev. Cancer 20, 89–106 (2020).
- Simpson, R. J. et al. Exercise and adrenergic regulation of immunity. *Brain Behav. Immun.* 97, 303–318 (2021).
- Campbell, J. P. & Turner, J. E. Debunking the myth of exercise-induced immune suppression: redefining the impact of exercise on immunological health across the lifespan. Front. Immunol. 9, 648 (2018).
- Campbell, J. P. et al. Acute exercise mobilises CD8+ T lymphocytes exhibiting an effector-memory phenotype. Brain Behav. Immun. 23, 767–775 (2009).
- 60. Bigley, A. B. et al. Acute exercise preferentially redeploys NK-cells with a highly differentiated phenotype and augments cytotoxicity against lymphoma and multiple myeloma target cells. *Brain Behav. Immun.* **39**, 160–171 (2014). This is a pioneering study showing that acute exercise may serve as a simple strategy to enrich the blood compartment of highly cytotoxic NK-cell subsets that can be harvested for clinical use (adoptive transfer immunotherapy).
- Batatinha, H. et al. Human lymphocytes mobilized with exercise have an anti-tumor transcriptomic profile and exert enhanced graft-versus-leukemia effects in xenogeneic mice. Front. Immunol. 14, 1067369 (2023).
- Zúňiga, T. M. et al. Acute exercise mobilizes NKT-like cells with a cytotoxic transcriptomic profile but does not augment the potency of cytokine-induced killer (CIK) cells. Front. Immunol. 13, 938106 (2022).
- 63. Turner, J. E. et al. Exercise-induced B cell mobilisation: preliminary evidence for an influx of immature cells into the bloodstream. *Physiol. Behav.* **164**, 376–382 (2016).
- Shephard, R. J. Adhesion molecules, catecholamines and leucocyte redistribution during and following exercise. Sports Med. 33, 261–284 (2003).
- 65. Graff, R. M. et al. β₂-Adrenergic receptor signaling mediates the preferential mobilization of differentiated subsets of CD8+ T-cells, NK-cells and non-classical monocytes in response to acute exercise in humans. *Brain Behav. Immun.* **74**, 143–153 (2018).
- Dimitrov, S., Lange, T. & Born, J. Selective mobilization of cytotoxic leukocytes by epinephrine. J. Immunol. 184, 503–511 (2010).
- Rehman, J. et al. Dynamic exercise leads to an increase in circulating ICAM-1: further evidence for adrenergic modulation of cell adhesion. *Brain Behav. Immun.* 11, 343–351 (1997).
- Goossens, G. H. et al. Short-term beta-adrenergic regulation of leptin, adiponectin and interleukin-6 secretion in vivo in lean and obese subjects. *Diabetes Obes. Metab.* 10, 1029–1038 (2008).
- Kruger, K., Lechtermann, A., Fobker, M., Volker, K. & Mooren, F. C. Exercise-induced redistribution of T lymphocytes is regulated by adrenergic mechanisms. *Brain Behav. Immun.* 22, 324–338 (2008).
 This is a study in rodents that elegantly shows (using fluorescent cell tracking) a

redistribution of T cells from the spleen to target organs (the lung, bone marrow and gut (Peyer's patches)) in the 24 hours after an acute exercise bout.

- Baker, F. L. et al. Systemic β-adrenergic receptor activation augments the ex vivo expansion and anti-tumor activity of Vy9Vδ2 T-cells. Front. Immunol. 10, 3082 (2020).
- Kruger, K. & Mooren, F. C. T cell homing and exercise. Exerc. Immunol. Rev. 13, 37–54 (2007).
- Kruger, K. et al. Apoptosis of T-cell subsets after acute high-intensity interval exercise. Med. Sci. Sports Exerc. 48, 2021–2029 (2016).
- Simpson, R. J. Aging, persistent viral infections, and immunosenescence: can exercise "make space"? Exerc. Sport Sci. Rev. 39, 23–33 (2011).
- Schenk, A. et al. Distinct distribution patterns of exercise-induced natural killer cell mobilization into the circulation and tumor tissue of patients with prostate cancer. *Am. J. Physiol. Cell Physiol.* **323**, C879–C884 (2022).
- Schauer, T., Djurhuus, S. S., Simonsen, C., Brasso, K. & Christensen, J. F. The effects of acute exercise and inflammation on immune function in early-stage prostate cancer. *Brain Behav. Immun. Health* 25, 100508 (2022).

- Djurhuus, S. S. et al. Effects of acute exercise training on tumor outcomes in men with localized prostate cancer: a randomized controlled trial. *Physiol. Rep.* 10, e15408 (2022).
- Djurhuus, S. S. et al. Exercise training to increase tumour natural killer-cell infiltration in men with localised prostate cancer: a randomised controlled trial. *BJU Int.* 131, 116–124 (2023).

This paper is a clinical trial showing that regular, intense exercise can increase NK cell infiltration in prostate tumours.

- Thienger, P. & Rubin, M. A. Prostate cancer hijacks the microenvironment. Nat. Cell Biol. 23, 3–5 (2021).
- 79. Martori, C. et al. Macrophages as a therapeutic target in metastatic prostate cancer: a way to overcome immunotherapy resistance? Cancers **14**, 440 (2022).
- Clifford, B. K., Kaakoush, N. O., Tedla, N., Goldstein, D. & Simar, D. The effect of exercise intensity on the inflammatory profile of cancer survivors: a randomized crossover study. *Eur. J. Clin. Invest.* 53, e13984 (2023).
- Valenzuela, P. L. et al. Exercise training and natural killer cells in cancer survivors: current evidence and research gaps based on a systematic review and meta-analysis. Sports Med. Open 8, 36 (2022).
- Coletta, A. M. et al. The impact of high-intensity interval exercise training on NK-cell function and circulating myokines for breast cancer prevention among women at high risk for breast cancer. Breast Cancer Res. Treat. 187, 407–416 (2021).
- Llavero, F. et al. Exercise training effects on natural killer cells: a preliminary proteomics and systems biology approach. *Exerc. Immunol. Rev.* 27, 125–141 (2021).
- 84. MacDonald, G. et al. A pilot study of high-intensity interval training in older adults with treatment naïve chronic lymphocytic leukemia. *Sci. Rep.* **11**, 3137 (2021).
- Toffoli, E. C. et al. Effects of physical exercise on natural killer cell activity during (neo) adjuvant chemotherapy: a randomized pilot study. *Physiol. Rep.* 9, e14919 (2021).
- 86. Mace, E. M. Phosphoinositide-3-kinase signaling in human natural killer cells: new insights from primary immunodeficiency. *Front. Immunol.* **9**, 445 (2018).
- Takahashi, N. et al. Tumor marker nucleoporin 88 kDa regulates nucleocytoplasmic transport of NF-kappaB. Biochem. Biophys. Res. Commun. 374, 424–430 (2008).
- Spielmann, G. et al. Aerobic fitness is associated with lower proportions of senescent blood T-cells in man. Brain Behav. Immun. 25, 1521–1529 (2011).
- Himbert, C. et al. Differences in the gut microbiome by physical activity and BMI among colorectal cancer patients. *Am. J. Cancer Res.* 12, 4789–4801 (2022).
- 90. O'Sullivan, O. et al. Exercise and the microbiota. *Gut Microbes* **6**, 131–136 (2015).
- Clarke, S. F. et al. Exercise and associated dietary extremes impact on gut microbial diversity. Gut 63, 1913–1920 (2014).
- Lozupone, C. A., Stombaugh, J. I., Gordon, J. I., Jansson, J. K. & Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* 489, 220–230 (2012).
- Wirbel, J. et al. Meta-analysis of fecal metagenomes reveals global microbial signatures that are specific for colorectal cancer. Nat. Med. 25, 679–689 (2019).
- Routy, B. et al. The gut microbiota influences anticancer immunosurveillance and general health. Nat. Rev. Clin. Oncol. 15, 382–396 (2018).
- Park, E. M. et al. Targeting the gut and tumor microbiota in cancer. Nat. Med. 28, 690–703 (2022).
- Gopalakrishnan, V. et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. Science 359, 97–103 (2018).
- Wang, B. et al. Synergetic inhibition of daidzein and regular exercise on breast cancer in bearing-4T1 mice by regulating NK cells and apoptosis pathway. *Life Sci.* 245, 117387 (2020).
- Dufresne, S. et al. Exercise training improves radiotherapy efficiency in a murine model of prostate cancer. FASEB J. 34, 4984–4996 (2020).
- Laskowski, T. J., Biederstädt, A. & Rezvani, K. Natural killer cells in antitumour adoptive cell immunotherapy. Nat. Rev. Cancer 22, 557–575 (2022).
- 100. Di Vito, C. et al. NK cells to cure cancer. Semin. Immunol. 41, 101272 (2019).
- Huntington, N. D., Cursons, J. & Rautela, J. The cancer-natural killer cell immunity cycle. Nat. Rev. Cancer 20, 437–454 (2020).
- López-Soto, A., Gonzalez, S., Smyth, M. J. & Galluzzi, L. Control of metastasis by NK cells. Cancer Cell 32, 135–154 (2017).
- Li, B., Jiang, Y., Li, G., Fisher, G. A. & Li, R. Natural killer cell and stroma abundance are independently prognostic and predict gastric cancer chemotherapy benefit. *JCI Insight* 5, e136570 (2020).
- Lee, H. et al. Integrated molecular and immunophenotypic analysis of NK cells in anti-PD-1 treated metastatic melanoma patients. *Oncoimmunology* 8, e1537581 (2019).
- Cursons, J. et al. A gene signature predicting natural killer cell infiltration and improved survival in melanoma patients. Cancer Immunol. Res. 7, 1162–1174 (2019).
- Buss, L. A. et al. Effects of exercise and anti-PD-1 on the tumour microenvironment. Immunol. Lett. 239, 60–71 (2021).
- Garritson, J. et al. Physical activity delays accumulation of immunosuppressive myeloid-derived suppressor cells. *PLoS ONE* 15, e0234548 (2020).
- Wennerberg, E. et al. Exercise reduces immune suppression and breast cancer progression in a preclinical model. Oncotarget 11, 452–461 (2020).
- Mengos, A. E., Gastineau, D. A. & Gustafson, M. P. The CD14⁺HLA-DR^{lo/neg} monocyte: an immunosuppressive phenotype that restrains responses to cancer immunotherapy. *Front. Immunol.* **10**, 1147 (2019).
- Kim, I. S. et al. Immuno-subtyping of breast cancer reveals distinct myeloid cell profiles and immunotherapy resistance mechanisms. *Nat. Cell Biol.* 21, 1113–1126 (2019).

- Weber, R. et al. Myeloid-derived suppressor cells hinder the anti-cancer activity of immune checkpoint inhibitors. Front. Immunol. 9, 1310 (2018).
- Martín-Ruiz, A. et al. Benefits of exercise and immunotherapy in a murine model of human non-small-cell lung carcinoma. Exerc. Immunol. Rev. 26, 100–115 (2020).
- Bay, M. L. et al. Voluntary wheel running can lead to modulation of immune checkpoint molecule expression. Acta Oncol. 59, 1447–1454 (2020).
- Denton, N. L., Chen, C. Y., Scott, T. R. & Cripe, T. P. Tumor-associated macrophages in oncolytic virotherapy: friend or foe? *Biomedicines* 4, 13 (2016).
- Rolny, C. et al. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through downregulation of PIGF. *Cancer Cell* 19, 31–44 (2011).
- Ruffell, D., Affara, N. I. & Coussens, L. M. Differential macrophage programming in the tumor microenvironment. *Trends Immunol.* 33, 119–126 (2012).
- Goh, J. et al. Exercise training in transgenic mice is associated with attenuation of early breast cancer growth in a dose-dependent manner. PLoS ONE 8, e80123 (2013).
- McClellan, J. L. et al. Exercise effects on polyp burden and immune markers in the ApcMin/+ mouse model of intestinal tumorigenesis. *Int. J. Oncol.* 45, 861–868 (2014).
- Ge, Z., Wu, S., Qi, Z. & Ding, S. Exercise modulates polarization of TAMs and expression of related immune checkpoints in mice with lung cancer. J. Cancer 13, 3297–3307 (2022).
- Lamkin, D. M. et al. Physical activity modulates mononuclear phagocytes in mammary tissue and inhibits tumor growth in mice. *PeerJ* 9, e10725 (2021).
- Castanedo-Rincón, C. et al. Combined exercise intervention in a mouse model of high-risk neuroblastoma: effects on physical, immune, tumor and clinical outcomes. *Exerc. Immunol. Rev.* 29, 86–110 (2023).
- Singh, G. & Singh, S. M. Role of host's antitumor immunity in exercise-dependent regression of murine T-cell lymphoma. *Comp. Immunol. Microbiol. Infect. Dis.* 28, 231–248 (2005).
- Broz, M. L. et al. Dissecting the tumor myeloid compartment reveals rare activating antigen-presenting cells critical for T cell immunity. *Cancer Cell* 26, 638–652 (2014).
- Ruffell, B. et al. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. Cancer Cell 26, 623–637 (2014).
- Layer, J. P. et al. Amplification of N-Myc is associated with a T-cell-poor microenvironment in metastatic neuroblastoma restraining interferon pathway activity and chemokine expression. Oncoimmunology 6, e1320626 (2017).
- Zafari, R., Razi, S. & Rezaei, N. The role of dendritic cells in neuroblastoma: implications for immunotherapy. *Immunobiology* 227, 152293 (2022).
- Martín-Ruiz, A. et al. Effects of anti-PD-1 immunotherapy on tumor regression: insights from a patient-derived xenograft model. Sci. Rep. 10, 7078 (2020).
- Hagar, A. et al. Endurance training slows breast tumor growth in mice by suppressing Treg cells recruitment to tumors. BMC Cancer 9, 536 (2019).
- Gomes-Santos, I. L. et al. Exercise training improves tumor control by increasing CD8+ T-cell infiltration via CXCR3 signaling and sensitizes breast cancer to immune checkpoint blockade. Cancer Immunol. Res. 9, 765–778 (2021).
- Rundqvist, H. et al. Cytotoxic T-cells mediate exercise-induced reductions in tumor growth. *eLife* 9, e59996 (2020).

Together with Gomes-Santos et al., this paper provides preclinical support for an increase in the anticancer effector function of CD8⁺ T cells with regular exercise.

- Feng, Q. et al. Lactate increases stemness of CD8+T cells to augment anti-tumor immunity. Nat. Commun. 13, 4981 (2022).
- Kaymak, I. et al. Carbon source availability drives nutrient utilization in CD8⁺ T cells. Cell Metab. 34, 1298–1311.e6 (2022).
- Barbieri, L. et al. Lactate exposure shapes the metabolic and transcriptomic profile of CD8+ T cells. Front. Immunol. 14, 1101433 (2023).
- Balachandran, V. P., Beatty, G. L. & Dougan, S. K. Broadening the impact of immunotherapy to pancreatic cancer: challenges and opportunities. *Gastroenterology* 156, 2056–2072 (2019).
- Bear, A. S., Vonderheide, R. H. & O'Hara, M. H. Challenges and opportunities for pancreatic cancer immunotherapy. *Cancer Cell* 38, 788–802 (2020).
- Gupta, P. et al. Comparison of three exercise interventions with and without gemcitabine treatment on pancreatic tumor growth in mice: no impact on tumor infiltrating lymphocytes. Front. Physiol. 13, 1039988 (2022).
- Laumont, C. M. & Nelson, B. H. B cells in the tumor microenvironment: multi-faceted organizers, regulators, and effectors of anti-tumor immunity. *Cancer Cell* 41, 466–489 (2023).
- Wieland, A. et al. Defining HPV-specific B cell responses in patients with head and neck cancer. Nature 597, 274–278 (2021).
- Cabrita, R. et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. Nature 577, 561–565 (2020).
- Petitprez, F. et al. B cells are associated with survival and immunotherapy response in sarcoma. Nature 577, 556–560 (2020).

- Zitvogel, L., Tesniere, A. & Kroemer, G. Cancer despite immunosurveillance: immunoselection and immunosubversion. Nat. Rev. Immunol. 10, 715–727 (2006).
- Schmitz, K. H. et al. Exercise is medicine in oncology: engaging clinicians to help patients move through cancer. *CA Cancer J. Clin.* 69, 468–484 (2019).
 Siversten, I. & Dahlstrom, A. W. Relation of muscular activity to carcinoma: a preliminary
- 144. Siversten, I. & Danistrom, A. W. Relation of muscular activity to carcinoma: a preliminary report. J. Cancer Res. 6, 365–378 (1921).
- 145. Rusch, H. P. & Kline, B. E. The effect of exercise on the growth of a mouse tumor. Cancer Res. 4, 116–118 (1944).
- Newton, G. Tumor susceptibility in rats: role of infantile manipulation and later exercise. Psychol. Rep. 16, 127–132 (1965).
- Deuster, P. A., Morrison, S. D. & Ahrens, R. A. Endurance exercise modifies cachexia of tumor growth in rats. *Med. Sci. Sports Exerc.* 17, 385–392 (1985).
- MacNeil, B. & Hoffman-Goetz, L. Exercise training and tumour metastasis in mice: influence of time of exercise onset. *Anticancer Res.* 13, 2085–2088 (1993).
- 149. Virchow, R. Cellular pathology. As based upon physiological and pathological histology. Lecture XVI – atheromatous affection of arteries. 1858. Nutr. Rev. 47, 23–25 (1989).
- 150. Starnes, C. O. Coley's toxins. *Nature* **360**, 23 (1992).
- Starnes, C. O. Coley's toxins in perspective. Nature 357, 11–12 (1992).
 MacNeil, B. & Hoffman-Goetz, L. Effect of exercise on natural cytotoxicity and pulmonary tumor metastases in mice. Med. Sci. Sports Exerc. 25, 922–928 (1993).
 This preclinical study investigates on whether regular exercise improves anticancer function (as assessed with splenic NK cell cytotoxic activity).
- Hutt, D. Feasibility of leukapheresis for CAR T-cell production in heavily pre-treated pediatric patients. *Transfus. Apher. Sci.* 59, 102769 (2020).
- Korell, F. et al. Current challenges in providing good leukapheresis products for manufacturing of CAR-T cells for patients with relapsed/refractory NHL or ALL. Cells 9, 1225 (2020).
- Tuazon, S. A. et al. Factors affecting lymphocyte collection efficiency for the manufacture of chimeric antigen receptor T cells in adults with B-cell malignancies. *Transfusion* 59, 1773–1780 (2019).
- Allen, E. S. et al. Autologous lymphapheresis for the production of chimeric antigen receptor T cells. *Transfusion* 57, 1133–1141 (2017).
- Hont, A. B. et al. The generation and application of antigen-specific T cell therapies for cancer and viral-associated disease. *Mol. Ther.* **30**, 2130–2152 (2022).
- LaVoy, E. C. et al. A single bout of dynamic exercise enhances the expansion of MAGE-A4 and PRAME-specific cytotoxic T-cells from healthy adults. *Exerc. Immunol. Rev.* 21, 144–153 (2015).

Acknowledgements

The authors are grateful to K. McCreath for helpful comments on the text. Research by A.L. and C.F.-L. in exercise and cancer is funded by the Wereld Kanker Onderzoek Fonds (WKOF), as part of the World Cancer Research Fund International grant programme (grant number IIG_FULL_2021_007), and the Spanish Ministry of Science and Innovation (Fondo de Investigaciones Sanitarias (FIS)) and Fondos FEDER (grant numbers P118/00139 and P120/00645), and European Union's Horizon 2020 research and innovation programme under grant agreement number 945153. Research by C.F.-L. is funded by a Miguel Servet postdoctoral contract granted by Instituto de Salud Carlos III (CP18/00034). Research by P.L.V. is funded by a Sara Borrell postdoctoral contract granted by Instituto de Salud Carlos III (CD21/00138).

Author contributions

A.L. wrote the first manuscript draft with the help of C.F.-L. All authors researched data for the article, contributed to the discussion of content, and also reviewed and edited the article in depth before submission. B.G.G. and C.F.-L. made the figures.

Competing interests

The authors declare no conflict of interest.

Additional information

Peer review information Nature Reviews Immunology thanks Bente Klarlund Pedersen, Karsten Krueger and John P. Campbell for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2023